COWELL REGENERATIVE SOLUTION

Inspire confidence through a comprehensive approach

• InnoGraft B

A xenograft composed of 100% bovine cancellous bone with 3-Dimensional structures that allow optimal cell attachment and blood penetration.



The WORLD'S FIRST E.rhBMP-2-based bone graft material that induces bone and cartilage formation as a retinoid mediator that plays a key role in osteoblast differentiation.

with the highest pharmacological standard of AATB.

COWELL REGENERATIVE SOLUTION

→INNO CaP

An osteoconductive resorbable synthetic bone graft material composed of 100% calcium phosphate to be progressively replaced by normal-structured bone in the healing period.





Osteoinductive Bone Graft rhBMP-2 + BCP/DCP



The world's first E.rhBMP-2 (E.Coli derived Recombinant Human Bone Morphogenetic Protein type 2), as a growth factor that induces bone and cartilage formation. It is a retinoid mediator that plays a key role in osteoblast differentiation.

Composition

COWELL BMP is bone graft material based on the E.rhBMP-2, developed for the first time in the world. It is supported by 10 years of clinical data and over 40 studies.
BCP/DCP as a carrier allows maintenance of space.

Features

- Primary closure for soft tissue regeneration is not required.
- Regenerates adherent gingiva.
- Simplifies challenging bone grafting and soft tissue regeneration.
- Acts directly on stem cells.
- Induces bone regeneration without infection in extraction socket.
- Contains 1mg of bone morphogenic protein per 1g of the particle (1g of autologous bone contains 2ng of bone morphogenic protein).

Experience innovation COVELL BMP

THE WORLD FIRST E.rhBMP-2-based bone graft, supported by **10 YEARS OF CLINICAL DATA AND 40+ STUDIES.**

rhBMP2

COWELL IMPLANT SYSTEM

COWELLMEDI HISTORY

REID

Development Background

Triad of Tissue Engineering



Autologous stem cell transplantation

- Less effective due to difficulty of the engraftment in early stage of tissue regeneration
- Cell cultivation causes enormous expense

However, Stem cell growth factors

- Effective in tissue regeneration for all vertebrates
- Even human growth factor is effective in both human and animals

Stem cell transplantation VS rhBMP-2





Stem cell transplantation





Mechanism of Action of COWELL BMP







Mesenchymal Stem cell

- 1. rhBMP-2 bonds with BMP-2 receptor of Stem cell to activate intracellular phosphorylating enzyme.
- BMP-2 of Stem cell and VEFG activates for protein synthesis and secretion.
 * VEGF : Vascular Endothelial Growth Factor



- 3. VEGF promotes cell growth by inducing angiogenesis to nourish Stem cell.
- BMP-2
- 4. BMP-2, activates cell division of surrounding Stem cell and promotes rapid proliferation.



5. Proliferated Stem cells, differentiate into various cells according to surrounding tissues.



6. Differentiated cells form neoplastic tissues and remodel them according to the surrounding environment.

Product Type

COWELL BMP (One vial)



• Dose and particle size of the COWELL BMP



* A vial of 0.1g can be used for less than one extraction socket, while 0.25g/0.5g can be used for maxillary sinus or for the wide bone defect area.



COWELL BMP Plus (Two vials)



• Dose and particle size of the COWELL BMP Plus.

BMP 0.1mg						
Product Code	BMP Dose	Particle Dose	Particle Size			
EBB0125	0.1mg	0.25g	0.41~1.0mm			
EBB0105	0.1mg	0.5g	0.41~1.0mm			
EBB1110	0.1mg	1g	0.41~1.0mm			
EBB1220	0.1mg	2g	0.41~1.0mm			

BMP 0.5mg

Product Code	BMP Dose	Particle Dose	Particle Size
EBB0525	0.5mg	0.25g	0.41~1.0mm
EBB0505	0.5mg	0.5g	0.41~1.0mm
EBB1150	0.5mg	1g	0.41~1.0mm
EBB1250	0.5mg	2g	0.41~1.0mm

BMP 2mg

Product Code	BMP Dose	Particle Dose	Particle Size
EBB2025	2mg	0.25g	0.41~1.0mm
EBB2050	2mg	0.5g	0.41~1.0mm
EBB2011	2mg	1g	0.41~1.0mm
EBB2012	2mg	2g	0.41~1.0mm

BMP 0.25mg

Product Code	BMP Dose	Particle Dose	Particle Size
EBB2525	0.25mg	0.25g	0.41~1.0mm
EBB2505	0.25mg	0.5g	0.41~1.0mm
EBB1125	0.25mg	1g	0.41~1.0mm
EBB1225	0.25mg	2g	0.41~1.0mm

BMP 1mg

Product Code	BMP Dose	Particle Dose	Particle Size
EBB1025	1mg	0.25g	0.41~1.0mm
EBB1050	1mg	0.5g	0.41~1.0mm
EBB1011	1mg	1g	0.41~1.0mm
EBB1012	1mg	2g	0.41~1.0mm





INNO GF Kit (Two vials + Saline Solution + Syringe)

• Dose and particle size of the INNO GF Kit.

BMP 0.1mg

Product Co	ode	BMP Dose	Particle Dose	Particle Size
IBB012	25	0.1mg	0.25g	0.41~1.0mm
IBB01	05	0.1mg	0.5g	0.41~1.0mm
IBB11	10	0.1mg	1g	0.41~1.0mm
IBB122	20	0.1mg	2g	0.41~1.0mm

BMP 0.5mg

Product Code	BMP Dose	Particle Dose	Particle Size
IBB0525	0.5mg	0.25g	0.41~1.0mm
IBB0505	0.5mg	0.5g	0.41~1.0mm
IBB1150	0.5mg	1g	0.41~1.0mm
IBB1250	0.5mg	2g	0.41~1.0mm

BMP 2mg

Product Code	BMP Dose	Particle Dose	Particle Size
IBB2025	2mg	0.25g	0.41~1.0mm
IBB2050	2mg	0.5g	0.41~1.0mm
IBB2011	2mg	1g	0.41~1.0mm
IBB2012	2mg	2g	0.41~1.0mm

BMP 0.25mg

Product Code	BMP Dose	Particle Dose	Particle Size
IBB2525	0.25mg	0.25g	0.41~1.0mm
IBB2505	0.25mg	0.5g	0.41~1.0mm
IBB1125	0.25mg	1g	0.41~1.0mm
IBB1225	0.25mg	2g	0.41~1.0mm

BMP 1mg

	Product Code	BMP Dose	Particle Dose	Particle Size
	IBB1025	1mg	0.25g	0.41~1.0mm
	IBB1050	1mg	0.5g	0.41~1.0mm
	IBB1011	1mg	1g	0.41~1.0mm
	IBB1012	1mg	2g	0.41~1.0mm



REID

User Guide COWELL BMP

A. Method I



a. Transfer DCP graft material (Vial $I\!\!I$).



c. Mix BMP solution with DCP or plus autogenic / allograft and, apply to the recipient site.

B. Method II



a. Transfer DCP graft material (Vial I) into a container.





b. Inject distilled water into vial ${\rm I\!I}$ with lyophilized rhBMP-2 power in it and mix with the powder.

d. Cover the defect area using a barrier membrane or

suture natural soft tissue without membrane.

b. Apply DCP into the recipient site and cover the defect area using a barrier membrane or suture natural soft tissue without membrane.



c. Inject distilled water into lyophilized rhBMP-2 powder (vial II).



d. Mix rhBMP-2 with distilled water (saline solution) and wait for 10 to 15 seconds before using.



e. Aspirate the mixture using a syringe.



f. Inject BMP solution through soft tissue until needle of syringe reaches bone.

C. Method III





c. Hydrate BMP-2 solution into membrane.



d. Apply BMP-2 solution socked membrane to damaged site.

Dose of distilled water to make the mixture (BMP-2 Solution)

BMP Dose	Distilled Water Dose	BMP Dose	Distilled Water Dose
0.1mg	0.1ml	2mg	1.6ml
0.25mg	0.2ml	5mg	4ml
0.5mg	0.4ml	10mg	8ml
1mg	0.8ml	20mg	16ml

Video

* Scan above QR code to watch videos of user guide of COWELL BMP

1. Mixture with bone graft material

Full dose of COWELL BMP

Excess leakage of COWELL BMP

Douse bone graft material immediately before the graft to minimize the time for rhBMP-2 protein to adsorb to bone graft calcium ingredient.









Collagen Plug



Bone matrix

2. Injection into bone graft site

1/2 dose of COWELL BMP

Moderate leakage of COWELL BMP

Even if the solution leaks out of the gingival after the injection, the effect is the same since the minimum effective drug dose has reached the stem cells.



General Syringe





Lidocaine Syringe



3. COWELL BMP coated implant

¹/₂ dose of COWELL BMP

Moderate leakage of COWELL BMP

The bone marrow stem cells are directly activated by placement of rhBMP-2 coated implant.



INNO Implant_1



INNO Implant_2



COWELL BMP 285

Safety of COWELL BMP

Q: Bone overgrowth by rhBMP-2?

A : rhBMP-2 is safe from bone overgrowth because Twist-2 is synthesized in Stem cells to stop cell division when bone formation period is completed.



Cell, Vol. 112, 169–180, January 24, 2003 European Journal of Endocrinology (2000) 142 9–21



Cell proliferation Cell differentiation

- \cdot Bonding to BMP-2 receptor
- · Signal pathway
- · Nuclear activation

VEGF, BMP Synthesis

Q : Correlation between cancer incidence and usual dose of rhBMP-2?

A : Generally, rhBMP-2 may be related to cancer incidence only when total dose is over 40mg. Countless research has proven that the safety standard dose is 4.2~8.4mg. COWELL BMP is supplied below the safety standard dose only.

(E.g. COWELLBMP 0.25g contains 0.25mg of rhBMP-2 which is 15 to 30 times lower than the safety standard.)



Q : Swelling occurrence after using rhBMP-2?

A : Relief incision may cause swelling due to angiogenesis proliferation in muscle but it is pain-free. Also, swelling is a transitional phenomenon and it is not a side effect.



Q : Seroma occurrence after using rhBMP-2?

A : After sinus lift surgery, excessive secretion of exudate during healing period may undertow in the grafted site of sealed maxillary sinus and develop into seroma but soon disappear. To limit the use to a maximum of 0.25 mg is safer rather than a high dose.



Effectiveness of COWELL BMP

Critically Defected Model

Bone Graft Type

Without rhBMP-2

With rhBMP-2







rhBMP-2 Coated Implant

Vertical Defect



Dehiscence Defect

Bone Graft



* Bone is safely formed without barrier membrane after rhBMP-2 bone graft, however, when use of general bone graft, barrier membrane is essential

rhBMP-2 Bone Graft



Effectiveness of COWELL BMP

Comparison with other materials

Both Calcium Pyrophosphate, CPP(Ca/P=1) and Biphasic Calcium Phosphate, BCP(Ca/P=1.55) are very effective for early osteoanagenesis. CPP, however, has higher absorption rate than BCP and is slightly more effective for osteoanagenesis.





There is no difference in the ratio of new bone generation.

However, Graft B forms hard fibrous tissue between particles and the COWELL BMP fills bone marrow tissue.

The Graft B received site has high resistance against drilling while the COWELL BMP has excellence in bone remodeling by bone.



Control ("Graft B")



COWELL BMP

CLINICAL CASE

Case 1.

Bone Regeneration and Gingival Improvement Using Bone Augmentation using COWELL BMP



Dr. Claudio Sotomayor Julio, D.D.S. Chille



① Pre-operative



③ 2 layers of membrane placement with COWELL BMP BCP powder



(2) INNO implant placement



4 COWELL BMP injection





(5) Post-operative



(7) 4 months healing period and removal of adhesive provisional tooth





8 2 weeks after connection surgery







Pre-operation (18.08.02)



Post-operation (18.08.02)



10 5 month after surgery : final rehabilitation



4 months (18. 12. 03)



1 year (19. 08. 06)

CLINICAL CASE

Case 2.

Bone regeneration in combination of rhBMP-2 and autogenous bone

62 years old, Female



Preoperative 2010. 04. 05



Postoperative 2010. 04. 05



10 months 2011.02.25



8 years 2019. 01. 18

CLINICAL CASE

Case 3. Staged implantation in healed ridge and extraction socket

63 years old, Male



Preoperative 2010. 04. 06



Postoperative 2010. 04. 30



9 months 2011.01.19



8 years 2019. 01. 08

Scientific Proofs of COWELL BMP's Effectiveness

- 1. Analysis of hydrolyzable polyethylene glycol hydrogels and deproteinized bone mineral as delivery systems for glycosylated and non-glycosylated bone morphogenetic protein-2. Acta Biomater. 2012 Jan;8(1):116-23.
- 2. Effects of rhBMP-2 Coating Tricalcium Phosphate on Socket Preservation in Dog Extraction Socket. Tissue Engineering and Regenerative Medicine, Vol. 5, No. 4~6, pp 637-642 (2008)
- 3. Effects of Polycaprolactone-Tricalcium Phosphate, Recombinant Human Bone Morphogenetic Protein-2 and Dog Mesenchymal Stem Cells on Bone Formation: Pilot Study in Dogs. Yonsei Med J 50(6): 825-831,(2009)
- The induction of bone formation in rat calvarial defects and subcutaneous tissues by recombinant human BMP-2, produced in Escherichia coli. Biomaterials 31 (2010) 3512–3519
- 5. Alveolar ridge augmentation using anodized implants coated with Escherichia coli–derived recombinant human bone morphogenetic protein 2.
 - Oral Surg Oral Med Oral Pathol Oral Radiol Endod. (2011) Jul;112(1):42-9 Bone formation of Escherichia coli expressed rbBMP-2 on absorbable collagen block in
- 6. Bone formation of Escherichia coli expressed rhBMP-2 on absorbable collagen block in rat calvarial defects. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2011;111:298-305
- Bone formation of block and particulated biphasic calcium phosphate lyophilized with Escherichia coli–derived recombinant human bone morphogenetic protein 2 in rat calvarial defects. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2011;112:298-306.
- 8. Induction of bone formation by Escherichia coli- expressed recombinant human bone morphogenetic protein-2 using block-type macroporous biphasic calcium phosphate in orthotopic and ectopic rat models. J Periodontal Res. (2011) Dec; 46(6):682-90.
- 9. Enhanced adipogenic differentiation and reduced collagen synthesis induced by human periodontal ligament stem cells might underlie the negative effect of recombinant human bone morphogenetic protein-2 on periodontal regeneration.
 - J Periodontal Res (2011); 46: 193-203
- 10. The Effects of rhBMP-2 Injection at Distraction Osteogenesis of Rats'Tibia. Tissue Engineering and Regenerative Medicine, Vol. 8, No. 2, pp 158-163 (2011).
- 11. Discontinuous Release of Bone Morphogenetic Protein-2 Loaded Within Interconnected Pores of Honeycomb-Like Polycaprolactone Scaffold Promotes Bone Healing in a Large Bone Defect of Rabbit Ulna. Tissue Eng Part A. 2011 Oct;17(19-20):2389-97.v
- The effect of immobilization of heparin and bone morphogenic protein-2 to bovine bone substitute on osteoblast-like cell's function.
 J Adv Prosthodont 2011: 3:145-51
- Multicenter, randomized clinical trial on the efficacy and safety of Escherichia coli-derived rhBMP-2 with β-Tricalcium phosphate and hydroxyapatite in human extraction sockets. J Adv Prosthodont 2011; 4:178-182
- 14. Effects of Anodized Implants Coated With Escherichia coli-Derived Recombinant Human Bone Morphogenetic Protein-2 on Osseointegration in Rabbits. Tissue Engineering and Regenerative Medicine, Vol. 8, No. 1, pp 62-68 (2011)
- 15. Novel analysis model for implant osseointegration using ectopic bone formation via the recombinant human bone morphogenetic protein-2/macroporous biphasic calcium phosphate block system in rats: a proof-of concept study.

J Periodontal Implant Sci 2012; 42:136-143

- 16. Effects of anodized implants coated with Escherichia coli-derived rhBMP-2 in beagle dogs. Int. J. Oral Maxillofac. Surg. 2012; 41: 1577–1584.
- 17. Bone formation of middle ear cavity using biphasic calcium phosphate lyophilized with Escherichia coli-derived recombinant human bone morphogenetic protein 2 using animal model. International Journal of Pediatric Otorhinolaryngology 77 (2013) 1430–1433
- 18. Bone formation and remodeling of three different dental implant surfaces with Escherichia coli-derived recombinant human bone morphogenetic protein 2 in a rabbit model. Int J Oral Maxillofac Implants, 2013; 28(2):424-30
- Recombinant Human Bone Morphogenetic Protein-2 Stimulates the Osteogenic Potential of the Schneiderian Membrane: A Histometric Analysis in Rabbits. Tissue Eng Part A. 2013 Sep;19(17-18):1994-2004
- 20. The effect of anodized implants coated with combined rhBMP-2 and recombinant human vascular endothelial growth factors on vertical bone regeneration in the marginal portion of the peri-implant. Oral Surg Oral Med Oral Pathol Oral Radiol 2013;115:e24-e31.
- 21. Sinus augmentation using BMP-2 in a bovine hydroxyapatite/collagen carrier in dogs. J Clin Periodontol 2014; 41: 86–93.
- 22. Low-Dose Recombinant Human Bone Morphogenetic Protein-2 to Enhance the Osteogenic Potential of the Schneiderian Membrane in the Early Healing Phase: In Vitro and In Vivo Studies. J Oral Maxillofac Surg 72:1480-1494, 2014
- 23. Prospective randomized, controlled trial of sinus grafting using Escherichiacoli-produced rhBMP-2 with a biphasic calcium phosphate carrier compared to deproteinized bovine bone. Clin Oral Implants Res. 2015 Dec;26(12):1361-8.
- 24. Controlled release of BMP-2 using a heparin-conjugated carrier system reduces in vivo adipose tissue formation.
 - J Biomed Mater Res A. 2015 Feb;103(2):545-54.
- 25. The efficacy of BMP-2 preloaded on bone substitute or hydrogel for bone regeneration at peri-implant defects in dogs.

Clin Oral Implants Res. 2015 Dec;26(12):1456-65.

- 26. Effect of rhBMP-2 Immobilized Anorganic Bovine Bone Matrix on Bone Regeneration. Int. J. Mol. Sci. 2015, 16, 16034-16052.
- Effects of rhBMP-2 on Sandblasted and Acid Etched Titanium Implant Surfaces on Bone Regeneration and Osseointegration: Spilt-Mouth Designed Pilot Study. Biomed Res Int. 2015; 2015:459393.
- 28. Comparison of collagen membrane and bone substitute as a carrier for rhBMP-2 in lateral onlay graft. Clin Oral Implants Res. 2015;26(1):e13-9.
- 29. Effects of BMP-2 Delivery in Calcium Phosphate Bone Graft Materials with Different Compositions on Bone Regeneration.
 - Materials 2016, 9, 954
- 30. Source and Carrier Effect on the Bioactivity of BMP Bio-Implants. Master of Science 2013. Sylvie Di Lullo 2013, Faculty of Dentistry, University of Toronto
- 31. Soft and hard tissue changes when socket preservation using rhBMP-2, PRP and Non-Resorbable dPTFE membrane.
 - Dental implant Journal: Vol. 3, May, 2014
- 32. The effect of rhBMP-2 bonegraft on infrabony defects. Dental implant Journal: Vol. 3, May, 2014

INNO-CaP Calcium Phosphate , Synthetic Bone Graft

Osteoconductive resorbable synthetic bone graft material

- INNO-CaP is an osteoconductive synthetic resorbable bone graft material consisting of Calcium Phosphate.
- INNO-CaP is completely resorbed and progressively replaced by normal-structured bone in the healing period.

Excellent Biocompatibility and Conductivity

• The characteristic biocompatibility and conductivity of the INNO-CaP represent the most safety.

Cell culture SEM images (14 days)



X1,000



X1,000



298 INNO-CaP

A porosity for new bone ingrowth

• The porosity promotes ingrowth of osteoblast, osteoclast, and growth factors.

Particle surface SEM image



Indications Sinus graft surgery

- For sinus graft, INNO-CaP is used alone or in combination with the other graft materials.
- Healing periods residual bone height.

residual bone height	less than 1mm	2~4mm	more than 4 mm
implant placement	post operation 9~12 months	post operation 6 months	simultaneous placement

GBR (Guided Bone Regeneration)

- Minimize the amount of autogenous bone.
- Sub-graft materials.
- Vertical and lateral augmentation.
- It is highly recommended to use with COWELL BMP.

Dose and Particle Size

Product Code	Particle Size	Particle Dose
IG1025	0.4~1.0mm	0.25g
IG1050		0.5g
IG1001		1g
IG1002		2g
IG1425	1.0~1.4mm	0.25g
IG1450		0.5g
IG1401		1g
IG1402		2g

CLINICAL CASE 1



#45 Implant placement



INNO CaP





POD 10 weeks



Pre-OP



Post-OP



POD 10 weeks



POD 1 year 6 months



Pre-OP



Severe defect



Vertical defect



Horizontal defect



INNO CaP



MegaDerm Plus



Healing period

POD 11 weeks



POD 11 weeks

POD 12 weeks



Pre-OP CT



Post-OP CT



POD 11 weeks



POD 1 year 9 months



POD 1 year 9 months

COWELLMEDI HISTORY

INNO OSS Allo

Allograft FDBA, Cortical 50% Cancellous 50%

Product Features

- This product is made up of human tissue from trusted donors whose gender, age, and medical history were checked to ensure that their tissue could be used safely.
- It is an ideal combination of 50% cortical powder and 50% cancellous powder for bone induction.
- The 50% cortical powder maintains the space of the transplanted area during the new bone formation due to the delayed absorption rate. [OsteoConduction]
- 50% cancellous powder is rich in minerals and collagen that promote cell adhesion, bone remodeling, and vascular re-formation. [OsteoInduction]
- To prevent cross-infection by a different donor, the process is done by a single donor.
- Under the higher-level pharmacological standards (medical criteria) of the American Association of Tissue Banks (AATB), we sampled, processed, and distributed the allograft tissue.
- We recommend use of this product with the COWELL BMP.
- INNO OSS Allo is classified as a MEDICAL DEVICE.

SEM Image

Specifications

Туре	Particle Size	Particle Dose
OSS3A	0.4 ~ 1.0mm	0.3g
OSS6A	0.4 ~ 1.0mm	0.6g

Method of Use



Remove the syringe's rubber cap.



Hydrate it in saline solution.



Turn and pull out the syringe cap to remove it.



Graft it in the desired area.



A Bone 100% fused to Natural Human Bone

- Fast blood penetration
- Super-hydrophilicity
- 3D structure
- Fast and easy to handle
- Maximizes bone fusion
- Mutually connected porosity
- Optimal cell attachment and blood absorption
- Stimulates the activity of osteoclasts and osteoblasts



COWELLMEDI HISTORY

50X Magnification

1000X Magnification

1500X Magnification

Fast and Perfect Blood Permeation by Super-Hydrophilicity

Safe & Trustable Material

- Made of 100% bovine cancellous bone.
- Cleansing more than 30 times to completely remove organic matter.
- Firmed bone formation as highly dense.
- 100% pure HA & 99.73% of bone crystallization.



Raw material



Graft test 1

Graft test 2

(New bone formation clearly observed around grafted bone site)

Specifications

Product Code	Particle Size	Volume
IGB2015	0.25~1.0mm	0.15g
IGB2025	0.25~1.0mm	0.25g
IGB2050	0.25~1.0mm	0.5g
IGB2100	0.25~1.0mm	1g

CLINICAL CASE 1



Fig 01. Preoperative radiograph.



Fig 02. Preoperative CBCT image. Sinusitis in bone sinus cavities.



Fig 03. Incision and flap elevation. Removal of granulation tissue.



Fig 04. Suction of pus from the sinus.

Fig 05. Bone grafting with InnoOss B. Resorbable membrane application.

Fig 06. Postoperative radiograph.

Fig 07. Preoperative CBCT image.

Fig 08. Postoperative radiograph at week 6. Fig 09. CBCT image of postoperative Final restoration delivery.

10 month.

CLINICAL CASE 2

Fig 01. Preoperative radiograph. 3months after extraction in lower left posterior.

Fig 02. Incision and flap elevation.

Fig 03. Implant placement on #35, 37.

Fig 04. Bone grafting with InnoOss B and InnoOss Allo.

Fig 05. Non-resorbable membrance application.

Fig 06. Suture.

Fig 07. Postoperative radiograph.

Fig 08. Postoperative CBCT image of #35(Lt), #37(Rt).

Fig 09. Clinical view of postoperative 2weeks.

Fig 10. Clinical view ofpostoperative 4months. Final restoration delivery.

Fig 11. Radiograph of postoperative 4 months.

Fig 12. 4 month postoperative CBCT image of #35(Lt), #37(Rt).

MEGA DERM PLUS Acellular Dermal Matrix

Product Features

- This product can carry out the functional blocks of the membrane (soft tissue penetration protection) due to its long absorption period, and has excellent manipulability.
- This product is produced under the stringent standards of the MFDS.
- The world's first E-Beam sterilization can induce safe and prompt engraftment.
- E-Beam is safe and can be effectively sterilized without destroying the collagen tissue structure.
- This product is the first in the world with the basement membrane layer removed (patent pending) to maximize the transplant engraftment rate.
- This shows the high engraftment rate after the transplant by maximizing the influx of fibroblasts and/or the neovascularization. (Patent Application No. 10-2012-0026616)

Application

- Mucogingival defect.
- Soft tissue formation around the implant area.
- Wide perforation in the Schneiderian membrane.

SEM Images (They have kept the collagen structure after the E-Beam sterilization.)

A. SEM (x200)

B. SEM (x20,000)

C. TEM (Transverse section)

D. TEM (Cross section)

Specifications

Product Code	Size	Thickness
D1520P	15x20mm	0.5~0.7mm
D1525P	15x25mm	0.5~0.7mm

MEGA DERM PLUS three-dimensional structure of the dermis

The world's first 'E-Beam' sterilization that does not destroy the collagen structure

Sacteria removalNormal collagenDestroyed collagenOchemical residue

InnoGenic Non-resorbable Membranes

InnoGenic Wifi-Mesh and InnoGenic PTFE-Mesh

• The InnoGenic Wifi-Mesh, PTFE-Mesh and Ti-Mesh are non-resorbable barrier membranes to be applied over intraoral defects, especially, tooth extraction and bone augmented sites. The InnoGenic Wifi-Mesh and PTFE-Mesh are made of proprietary 100% PTFE, the polytetrafluoroethylene (teflon) sheet which is a biologically inactive and tissue compatible material and the InnoGenic Wifi-Mesh is reinforced with titanium frames (Titanium Gr II, ASTM F 67) embedded between two layers of PTFE sheets.

InnoGenic Wifi-Mesh

> Packing unit: 1ea

Product Code	Size	Thickness
BTP1424AA	14X24	0.25
BTP1424AB	14X24	0.25
BTP1525BB	15X25	0.25
BTP1725CA	17X25	0.25
BTP1725CA12	17X25	0.25
BTP2030AB	20X30	0.25
BTP2030AB12	20X30	0.25
BTP2530AB	25X30	0.25
BTP2530AB15	25X30	0.25
BTP3040AB	30X40	0.25
BTP3040AB15	30X40	0.25

(00)

BTP1424AA

BTP1424AB

BTP1525BB

BTP1725CA / BTP1725CA12

* Titanium material is the same

REID

COWELL EXPERT INSTRUMENTS

Clinical Case using the Wifi-Mesh

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 Periodontitis with local osteomyelitis of #45 & 47

N

Bone graft using INNO-OSS Allo

 Shielding soft tissue penetration using Wifi-Mesh

Removal of Wifi-Mesh

 Dense periosteum layer has been formed

InnoGenic PTFE-Mesh

> Packing unit: 5ea

Product Code	Size	Thickness
TS 24301	24 x 30	0.1

Features

- Non-resorbable: Made of 100% non-resorbable material for users to modulate healing period.
- Non-porous (0.0 μm) + Open Membrane Sheet Technique: Prevention of infection or other possible defects caused

from passage or integration of bacteria through the porosity of plaster and it even allows to application of the Open Membrane Sheet

Technique

- Prevention of Displacement: Not only being sutured along with gingiva but also being fixed with components from the InnoGenic GBR Kit to prevent displacement of the product.
- **Close to Transparency:** Observation of the healing of the underlying tissue through almost transparent PTFE surface allows more predictable result and helps determine removal time easier.
- Easy to be Customized: Easy to modify the shape according to shape and dimension of the defect.
- Easy to be Removed : Put a hook in the hole of the titanium frame of the InnoGenic Wifi-Mesh and in any center part of the InnoGenic PTFE-Mesh and remove.

Indications

CLINICAL APPLICATION Wifi-Mesh

Case 1

Pre-op

Implant placement

Implant placement

Clinical occlusal view of #45 and #46 showed severe bone defects.

Buccal bone graft technique with Wifi-mesh of #45

Wifi-Mesh trimming

COWELLMEDI HISTORY

REID

Wifi-Meshes were applied to the defect.

Open membrane technique in extraction socket of #46

CLINICAL APPLICATION Wifi-Mesh

Case 2 _ Dr. Hoyeol Jang

Pre-OP panorama

Occlusal view of the bone defect

Flap reflection

Drilling

Implant placement of #43, 44, 45 & 46

Wifi-Mesh

Wifi-mesh preparation *It must be bent to form a shape, and If it is bent incompletely, it can spread inside the gingiva.

Wifi-mesh placement

Implant placement of #33, 34, 35 & 36

Releasing incision

Bone graft

Wifi-mesh preparation

Wifi-Mesh placement

Membrane holding suture

Primary suture

Post OP panorama

CLINICAL APPLICATION Wifi-Mesh

CT scan images after GBR shows significant amount of alveolar bone regeneration.

2 months after the 1st surgery

2nd surgery and Wifi-Mesh removal

The Wifi-mesh was easily removed.

The defect area was fully filled with the new bone.

Membrane removal

Installation of healing abutments

Both horizontal and vertical bone regeneration was noticed clinically.

Incision of #43 and 44

Uncovering surgery of Lower jaw

2nd OP panorama

POD 3 months Temporary loading

InnoGenic Non-resorbable Membranes 313

CLINICAL APPLICATION PTFE-Mesh

Case 1

Open membrane technique and immediate implant placement in maxillary molars

The maxillary molars were extracted. The PTFE-Mesh was covered over the bone graft of socket preservation and implants.

3 weeks. 3 weeks after the graft operation, the PTFE-Mesh was removed. The new keratinized gingiva was regenerated on the bone graft particles.

4 months. 4 months after the graft operation, the keratinized gingiva was regenerated in the defect of socket.

At visit.

Surgery.

6 months.

32 months.

After 6 months of implant placement, the splinted crown was placed. There was no loss of marginal bone at the 32 months follow-up visit.

As result, the immediate implant placement and the open membrane technique with socket bone graft could make the new keratinized gingiva.

CLINICAL APPLICATION PTFE-Mesh

Case 2

Lateral bone graft with immediate implant placement in mandibular molars

Lateral bone graft with implant placement was done in mandibular 1st molar.

The extraction sockets of 2nd molar and 2nd premolar were grafted with the open membrane technique.

3 weeks after the graft operation, the PTFE-Mesh was removed. The new keratinized gingiva was regenerated on the bone graft particles.

3 months after the graft operation, the keratinized gingiva was regenerated in the defect of socket.

At visit.

4 months.

Lateral bone graft.

During healing period, the crestal bone level was decreased in the site of lateral bone graft. From 4 months to 31 months of follow-up visit, there was no the loss of marginal bone. As result, lateral bone graft with implant placement and open membrane technique in extraction socket could make the new keratinized gingiva.

CLINICAL APPLICATION PTFE-Mesh

Case 3

Socket preservation with immediate implant placement in mandibular premolars

Socket bone graft with implant placement was done in the buccal wall defect of mandibular premolars. The extraction sockets of premolars were grafted with the open membrane technique.

3 weeks after the graft operation, the PTFE-Mesh was removed. The new keratinized gingiva was regenerated on the bone graft particles.

____3 months after the graft operation, the keratinized gingiva was regenerated ______ in the defect of socket, and the splinted crown was placed.

At visit.

15 months.

Surgery.

28 months.

3 months.

28 months of follow-up visit, there was no the loss of marginal bone.

As result, the open membrane technique with implant placement in he buccal wall defect of premolars could make the new keratinized gingiva.